

JLP PHLEBOTOMY PRACTICE EXAM (SAMPLE)

STUDY GUIDE



SAMPLE STUDY GUIDE WITH LIMITED QUESTIONS

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THE FULL VERSION STUDY GUIDE

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Introduction

Preparing for a certification exam can feel overwhelming, but with the right tools, it becomes an opportunity to build confidence, sharpen your skills, and move one step closer to your goals. At Examzify, we believe that effective exam preparation isn't just about memorization, it's about understanding the material, identifying knowledge gaps, and building the test-taking strategies that lead to success.

This guide was designed to help you do exactly that.

Whether you're preparing for a licensing exam, professional certification, or entry-level qualification, this book offers structured practice to reinforce key concepts. You'll find a wide range of multiple-choice questions, each followed by clear explanations to help you understand not just the right answer, but why it's correct.

The content in this guide is based on real-world exam objectives and aligned with the types of questions and topics commonly found on official tests. It's ideal for learners who want to:

- Practice answering questions under realistic conditions,
- Improve accuracy and speed,
- Review explanations to strengthen weak areas, and
- Approach the exam with greater confidence.

We recommend using this book not as a stand-alone study tool, but alongside other resources like flashcards, textbooks, or hands-on training. For best results, we recommend working through each question, reflecting on the explanation provided, and revisiting the topics that challenge you most.

Remember: successful test preparation isn't about getting every question right the first time, it's about learning from your mistakes and improving over time. Stay focused, trust the process, and know that every page you turn brings you closer to success.

Let's begin.

How to Use This Guide

This guide is designed to help you study more effectively and approach your exam with confidence. Whether you're reviewing for the first time or doing a final refresh, here's how to get the most out of your Examzify study guide:

1. Start with a Diagnostic Review

Skim through the questions to get a sense of what you know and what you need to focus on. Your goal is to identify knowledge gaps early.

2. Study in Short, Focused Sessions

Break your study time into manageable blocks (e.g. 30 – 45 minutes). Review a handful of questions, reflect on the explanations.

3. Learn from the Explanations

After answering a question, always read the explanation, even if you got it right. It reinforces key points, corrects misunderstandings, and teaches subtle distinctions between similar answers.

4. Track Your Progress

Use bookmarks or notes (if reading digitally) to mark difficult questions. Revisit these regularly and track improvements over time.

5. Simulate the Real Exam

Once you're comfortable, try taking a full set of questions without pausing. Set a timer and simulate test-day conditions to build confidence and time management skills.

6. Repeat and Review

Don't just study once, repetition builds retention. Re-attempt questions after a few days and revisit explanations to reinforce learning. Pair this guide with other Examzify tools like flashcards, and digital practice tests to strengthen your preparation across formats.

There's no single right way to study, but consistent, thoughtful effort always wins. Use this guide flexibly, adapt the tips above to fit your pace and learning style. You've got this!

Questions

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- 1. Which additive is required in the tube collected first for certain blood tests?**
 - A. Citrate
 - B. EDTA
 - C. Heparin
 - D. Sodium fluoride

- 2. Which document provides the phlebotomist with necessary information for laboratory specimen collection?**
 - A. Patient's medical history.
 - B. Billing statement.
 - C. Hospital protocol binder.
 - D. Laboratory requisition form.

- 3. For blood spot collection in newborns, what must be done before testing?**
 - A. The specimen must be completely dried before processing
 - B. Refrigerate immediately
 - C. Freeze before processing
 - D. Use a wet sample

- 4. What should be placed into an ice bath if processing is delayed for more than 30 minutes?**
 - A. N95 Respirator Mask
 - B. Seminal
 - C. Lactic Acid
 - D. Mask

- 5. Therapeutic Drug Monitoring involves which process?**
 - A. A process of checking liver enzymes
 - B. A process where a phlebotomist asks about the timing of a patient's last dose of medication.
 - C. A process of measuring glucose levels
 - D. A process of measuring electrolyte balance

- 6. Which collection tube is used for samples requiring anticoagulation to preserve cells for smears?**
- A. EDTA Tube
 - B. Heparin Tube
 - C. Citrate Tube
 - D. Plain Serum Tube
- 7. Implied consent is best described as which of the following?**
- A. Consent that is assumed in emergency situations where a patient is unable to provide explicit consent.
 - B. Written consent required for every procedure.
 - C. Consent from a family member is always sufficient.
 - D. Consent is never needed for emergency procedures.
- 8. Using tubes that have passed their expiration date may result in which of the following?**
- A. It improves sample stability.
 - B. Using tubes that have passed their expiration date may result in loss of vacuum.
 - C. It has no impact on specimen quality.
 - D. Expired tubes release stronger vacuum.
- 9. The minimum required number of unique patient identifiers a phlebotomist should use prior to a blood draw.**
- A. One
 - B. Two
 - C. Three
 - D. Four
- 10. Which site is preferred for dermal sampling in toddlers?**
- A. Non-dominant third finger
 - B. Dominant index finger
 - C. Heel
 - D. Palm

Answers

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1. A
2. D
3. A
4. C
5. B
6. A
7. A
8. B
9. B
10. A

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Explanations

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1. Which additive is required in the tube collected first for certain blood tests?

- A. Citrate**
- B. EDTA
- C. Heparin
- D. Sodium fluoride

Coagulation testing requires plasma that stays anticoagulated and free from substances that could prematurely affect clotting times. Citrate achieves this by binding calcium, a necessary factor in the coagulation cascade, effectively preventing clot formation. Collecting the first tube with citrate preserves the proper blood-to-anticoagulant ratio (blue-top tube) and minimizes contamination from other additives in later tubes, which is crucial for accurate PT and aPTT results. If a different additive were drawn first or if subsequent tubes contaminated the sample, calcium availability and coagulation factor activity could be altered, leading to inaccurate measurements. Other common additives serve different purposes—EDTA is used for CBCs, heparin for certain plasma tests, and fluoride for glucose measurement—so they're not suited to be the first tube for coagulation studies.

2. Which document provides the phlebotomist with necessary information for laboratory specimen collection?

- A. Patient's medical history.
- B. Billing statement.
- C. Hospital protocol binder.
- D. Laboratory requisition form.**

The document that guides a phlebotomist in specimen collection is the laboratory requisition form. It provides the official test orders and the precise collection details needed for each specimen: which tests are requested, the correct specimen type and tubes to use, any timing or fasting instructions, special handling or transport requirements, and patient identifiers to ensure proper labeling. This form connects the patient, the clinician, and the laboratory, ensuring that the right tests are performed on the correct patient and that results are associated with the right record. Other documents may offer context (like medical history) or general procedures (such as a hospital protocol binder), but they don't supply the specific, test-by-test instructions required for collecting and labeling specimens accurately.

3. For blood spot collection in newborns, what must be done before testing?

- A. The specimen must be completely dried before processing**
- B. Refrigerate immediately
- C. Freeze before processing
- D. Use a wet sample

Drying the blood spot completely before testing is essential. Dried spots on filter paper are stable and ready for reliable extraction of analytes; moisture left in the sample can cause diffusion, dilute or degrade components, promote contamination, and lead to inaccurate test results. After collection, allow the spot to air-dry thoroughly at room temperature on a clean surface away from heat and humidity, following the program's drying time, before proceeding with testing. Refrigerating or freezing before drying isn't part of normal pre-test handling, and a wet sample isn't acceptable.

4. What should be placed into an ice bath if processing is delayed for more than 30 minutes?

- A. N95 Respirator Mask
- B. Seminal
- C. Lactic Acid**
- D. Mask

When certain blood analytes are being measured, especially lactate, they can change quickly after collection due to ongoing cellular metabolism. Lactic acid increases if the sample isn't kept cold, because red blood cells and other cells keep glycolysis going. Putting the specimen in an ice bath slows enzymes and metabolic activity, helping preserve the in vivo lactate level until the sample can be processed. The other options don't relate to preserving a blood specimen for lactate testing, so they aren't appropriate choices in this context.

5. Therapeutic Drug Monitoring involves which process?

- A. A process of checking liver enzymes
- B. A process where a phlebotomist asks about the timing of a patient's last dose of medication.**
- C. A process of measuring glucose levels
- D. A process of measuring electrolyte balance

Therapeutic Drug Monitoring centers on obtaining a blood concentration of a drug at a specific time after dosing to guide dose adjustments and keep levels within a safe, effective range. The step where a phlebotomist asks about when the patient last took the medication is essential because the interpretation of the drug level depends on timing relative to the last dose. Drug levels rise after administration and fall as the body eliminates the drug, so knowing the exact timing helps determine whether the measured concentration reflects a trough, peak, or an in-between level and whether dose adjustments are needed. The other listed tasks—checking liver enzymes, measuring glucose, and assessing electrolyte balance—are important lab tests in their own right but are not part of the drug level monitoring process.

6. Which collection tube is used for samples requiring anticoagulation to preserve cells for smears?

- A. EDTA Tube**
- B. Heparin Tube
- C. Citrate Tube
- D. Plain Serum Tube

Preserving cell morphology while preventing clotting is essential for accurate smear evaluation. EDTA chelates calcium, stopping the coagulation cascade, so the blood remains fluid and the cells stay intact. This maintains the true shapes and staining properties of the cells, which is crucial for a good peripheral blood smear. Other options either lack an anticoagulant (plain serum) or use anticoagulants that can alter cell appearance or interfere with staining (citrate or heparin), making them less ideal for smears. So, an EDTA tube is the best choice for samples requiring anticoagulation to preserve cells for smears.

7. Implied consent is best described as which of the following?

- A. Consent that is assumed in emergency situations where a patient is unable to provide explicit consent.**
- B. Written consent required for every procedure.
- C. Consent from a family member is always sufficient.
- D. Consent is never needed for emergency procedures.

Implied consent exists to cover situations where a patient cannot communicate and delaying care would put them at risk. In emergencies, healthcare providers proceed with treatment that is necessary to prevent death or serious harm because the patient would likely consent to life-saving measures if able. This focuses on a reasonable belief of consent in urgent care, not on written forms or on always relying on family authorization. Written consent isn't required for every procedure, especially when time is critical. A family member's consent isn't automatically sufficient, and surrogate decision-making follows legal rules when the patient lacks capacity. Once the patient regains capacity or an advance directive or appropriate surrogate is available, consent should be reviewed and updated as needed.

8. Using tubes that have passed their expiration date may result in which of the following?

- A. It improves sample stability.
- B. Using tubes that have passed their expiration date may result in loss of vacuum.**
- C. It has no impact on specimen quality.
- D. Expired tubes release stronger vacuum.

The main idea is that evacuated blood collection tubes rely on a sealed vacuum to pull a precise amount of blood and to maintain the correct blood-to-additive ratio. Over time, the vacuum can degrade as the stopper or seal ages, or gas can diffuse through, causing a loss of vacuum. When the vacuum is reduced, the tube may draw too little blood or fill inconsistently, which can compromise test accuracy and sample quality. That's why using tubes past their expiration date may result in loss of vacuum. The other options aren't correct because expired tubes don't improve stability, don't have no impact, and don't create a stronger vacuum.

9. The minimum required number of unique patient identifiers a phlebotomist should use prior to a blood draw.

- A. One
- B. Two**
- C. Three
- D. Four

Verifying two unique patient identifiers before collecting a blood sample is essential to ensure the specimen goes with the right patient. Using two identifiers provides redundancy, so if one piece of information is incorrect or misread, there's still a check against the correct patient. Common pairs include full name and date of birth or a patient ID number and name. Relying on a single identifier leaves more room for mix-ups, especially with people who share names or in busy settings. While some high-risk situations might call for extra checks, two identifiers are the minimum standard for routine phlebotomy, and you should always confirm them against the patient's armband and the lab order before drawing and labeling.

10. Which site is preferred for dermal sampling in toddlers?

A. Non-dominant third finger

B. Dominant index finger

C. Heel

D. Palm

For dermal sampling in toddlers, the aim is to get a reliable blood sample with the least discomfort and quickest healing. The best site is the non-dominant ring finger. This choice offers a stable, easily accessible puncture point with a healthy capillary network, which helps yield an adequate sample with minimal tissue trauma. Using the non-dominant hand also reduces interference with the child's activities and minimizes pain on the hand you use most during daily tasks, making the procedure smoother for both the child and caregiver. Other sites are less favorable in toddlers. The heel is typically reserved for newborns and very young babies because heel punctures can be more painful and carry higher risk of injury and infection as children grow. The palm has thicker skin and less predictable capillary flow, making it harder to obtain a good sample and increasing discomfort. The dominant finger is avoided to lessen the child's overall pain and to prevent unnecessary pressure on the most frequently used hand. In practice, warming the hand and using a gentle puncture on the side of the fingertip of the non-dominant ring finger helps ensure a good blood flow and comfortable experience for the child.

Next Steps

Congratulations on reaching the final section of this guide. You've taken a meaningful step toward passing your certification exam and advancing your career.

As you continue preparing, remember that consistent practice, review, and self-reflection are key to success. Make time to revisit difficult topics, simulate exam conditions, and track your progress along the way.

If you need help, have suggestions, or want to share feedback, we'd love to hear from you. Reach out to our team at hello@examzify.com.

Or visit your dedicated course page for more study tools and resources:

<https://jlpplbotomy.examzify.com>

We wish you the very best on your exam journey. You've got this!

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