

ASCP TECHNOLOGIST IN CHEMISTRY (C) PRACTICE EXAM (SAMPLE)

STUDY GUIDE



**SAMPLE STUDY GUIDE
WITH LIMITED QUESTIONS**

**VISIT US ONLINE FOR
THE FULL VERSION STUDY GUIDE**

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Introduction

Preparing for a certification exam can feel overwhelming, but with the right tools, it becomes an opportunity to build confidence, sharpen your skills, and move one step closer to your goals. At Examzify, we believe that effective exam preparation isn't just about memorization, it's about understanding the material, identifying knowledge gaps, and building the test-taking strategies that lead to success.

This guide was designed to help you do exactly that.

Whether you're preparing for a licensing exam, professional certification, or entry-level qualification, this book offers structured practice to reinforce key concepts. You'll find a wide range of multiple-choice questions, each followed by clear explanations to help you understand not just the right answer, but why it's correct.

The content in this guide is based on real-world exam objectives and aligned with the types of questions and topics commonly found on official tests. It's ideal for learners who want to:

- Practice answering questions under realistic conditions,
- Improve accuracy and speed,
- Review explanations to strengthen weak areas, and
- Approach the exam with greater confidence.

We recommend using this book not as a stand-alone study tool, but alongside other resources like flashcards, textbooks, or hands-on training. For best results, we recommend working through each question, reflecting on the explanation provided, and revisiting the topics that challenge you most.

Remember: successful test preparation isn't about getting every question right the first time, it's about learning from your mistakes and improving over time. Stay focused, trust the process, and know that every page you turn brings you closer to success.

Let's begin.

How to Use This Guide

This guide is designed to help you study more effectively and approach your exam with confidence. Whether you're reviewing for the first time or doing a final refresh, here's how to get the most out of your Examzify study guide:

1. Start with a Diagnostic Review

Skim through the questions to get a sense of what you know and what you need to focus on. Your goal is to identify knowledge gaps early.

2. Study in Short, Focused Sessions

Break your study time into manageable blocks (e.g. 30 – 45 minutes). Review a handful of questions, reflect on the explanations.

3. Learn from the Explanations

After answering a question, always read the explanation, even if you got it right. It reinforces key points, corrects misunderstandings, and teaches subtle distinctions between similar answers.

4. Track Your Progress

Use bookmarks or notes (if reading digitally) to mark difficult questions. Revisit these regularly and track improvements over time.

5. Simulate the Real Exam

Once you're comfortable, try taking a full set of questions without pausing. Set a timer and simulate test-day conditions to build confidence and time management skills.

6. Repeat and Review

Don't just study once, repetition builds retention. Re-attempt questions after a few days and revisit explanations to reinforce learning. Pair this guide with other Examzify tools like flashcards, and digital practice tests to strengthen your preparation across formats.

There's no single right way to study, but consistent, thoughtful effort always wins. Use this guide flexibly, adapt the tips above to fit your pace and learning style. You've got this!

Questions

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- 1. Other than pancreatitis, in which condition can lipase be increased?**
 - A. Hypertension
 - B. Peptic ulcers
 - C. Gastroesophageal reflux
 - D. Allergic reactions

- 2. What reagent is specifically used in the direct HDL cholesterol method?**
 - A. Sulfated cyclodextrin
 - B. 4-Aminoantipyrine
 - C. Potassium bromide
 - D. Glycerol kinase

- 3. What is the primary source of plasma ammonia?**
 - A. It is produced in the liver from breakdown of fats
 - B. It is produced in the intestines from the breakdown of proteins by bacterial enzymes
 - C. It is produced in the kidneys from amino acid metabolism
 - D. It is produced in the muscles during physical activity

- 4. What aspect of polyacrylamide is notable concerning safety when preparing gels?**
 - A. It is non-toxic
 - B. It requires a protective barrier
 - C. It is flammable
 - D. Acrylamide is potentially carcinogenic

- 5. What is required to form a holoenzyme?**
 - A. Apoenzyme plus cofactor
 - B. Apoenzyme plus substrate
 - C. Apoenzyme plus prosthetic group
 - D. Apoenzyme plus reaction product

- 6. How are most enzymes monitored in clinical settings?**
- A. By measuring the rate of temperature change
 - B. By observing color changes in the reaction
 - C. By monitoring the rate of absorbance change
 - D. By assessing pH fluctuations
- 7. What is the order of ALP isoenzyme electrophoresis migration from slowest to fastest?**
- A. Renal - Intestinal - Bone - Liver - Placental
 - B. Renal - Bone - Intestinal - Liver - Placental
 - C. Renal - Intestinal - Placental - Bone - Liver
 - D. Renal - Intestinal - Bone - Placental - Liver
- 8. Amylase is classified as what type of enzyme?**
- A. Oxidase
 - B. Hydrolase
 - C. Transferase
 - D. Ligase
- 9. In a lipid panel, which two tests are least affected by fasting?**
- A. LDL cholesterol and triglycerides
 - B. Total cholesterol and HDL cholesterol
 - C. Triglycerides and HDL cholesterol
 - D. LDL cholesterol and total cholesterol
- 10. Which laboratory phenomenon can affect uric acid testing results significantly?**
- A. Specimen hemolysis
 - B. Refrigeration of the sample
 - C. Delayed serum separation
 - D. Prolonged exposure to light

Answers

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1. B
2. A
3. B
4. D
5. C
6. C
7. D
8. B
9. B
10. A

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Explanations

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1. Other than pancreatitis, in which condition can lipase be increased?

- A. Hypertension
- B. Peptic ulcers**
- C. Gastroesophageal reflux
- D. Allergic reactions

Lipase is an enzyme primarily produced by the pancreas that plays a crucial role in the digestion of fats. While most commonly associated with pancreatitis, lipase can also be elevated in other conditions, particularly in situations where there is pancreatic damage or dysfunction. Peptic ulcers, which are sores that develop on the lining of the stomach or the first part of the small intestine, can cause irritation and inflammation that may affect nearby structures, including the pancreas. In cases of complicated ulcers or when there is a concern of associated conditions such as perforation or gastritis, the pancreas can become reactive or stressed, leading to the release of lipase into the bloodstream. Conditions like hypertension, gastroesophageal reflux, and allergic reactions do not have a direct association with increased lipase levels. Hypertension primarily affects the cardiovascular system, gastroesophageal reflux is related to esophageal irritation and does not typically involve pancreatic enzyme changes, and allergic reactions usually result in histamine-mediated responses rather than pancreatic enzyme alterations. Therefore, among these options, peptic ulcers are the most pertinent condition where lipase levels can rise outside of pancreatitis.

2. What reagent is specifically used in the direct HDL cholesterol method?

- A. Sulfated cyclodextrin**
- B. 4-Aminoantipyrine
- C. Potassium bromide
- D. Glycerol kinase

The direct HDL cholesterol method specifically utilizes sulfated cyclodextrin as a reagent. This compound plays a critical role in the selective separation of HDL cholesterol from other lipoproteins in the sample. By forming a complex with other lipoproteins, sulfated cyclodextrin allows for the accurate measurement of HDL cholesterol levels without the need for prior precipitation methods that can be cumbersome and time-consuming. In this context, the function of sulfated cyclodextrin exemplifies its utility in enhancing specificity and sensitivity in lipid assays. By promoting the solubilization of HDL, the reagent ensures that the subsequent enzymatic reactions can effectively target only the HDL fraction, leading to more reliable and reproducible results in laboratory analysis.

3. What is the primary source of plasma ammonia?

- A. It is produced in the liver from breakdown of fats
- B. It is produced in the intestines from the breakdown of proteins by bacterial enzymes**
- C. It is produced in the kidneys from amino acid metabolism
- D. It is produced in the muscles during physical activity

The primary source of plasma ammonia is indeed produced in the intestines from the breakdown of proteins by bacterial enzymes. In the human digestive system, dietary proteins are broken down into amino acids and other nitrogenous compounds. The bacteria present in the intestines further metabolize these proteins, resulting in the production of ammonia as a byproduct, which is then absorbed into the bloodstream. This ammonia can contribute to various metabolic processes and is particularly important to consider in clinical settings, as elevated levels might indicate liver dysfunction or issues with the urea cycle. The liver typically converts ammonia into urea, which is then excreted by the kidneys. Therefore, ammonia levels in the blood can provide valuable insights into both hepatic function and protein metabolism. The other sources mentioned—such as the liver, kidneys, and muscles—do participate in ammonia production, but the contribution from intestinal bacteria is the most significant in terms of the primary source that influences plasma ammonia concentrations in a healthy individual.

4. What aspect of polyacrylamide is notable concerning safety when preparing gels?

- A. It is non-toxic
- B. It requires a protective barrier
- C. It is flammable
- D. Acrylamide is potentially carcinogenic**

Acrylamide, a key component in the preparation of polyacrylamide gels, is classified as a potential carcinogen. This is significant because it poses a health risk during the preparation of gels, particularly if inhaled or if it comes into contact with the skin. Workers should be aware of this risk and should use appropriate safety measures to reduce exposure, such as working in a fume hood, wearing gloves, and using protective eyewear. The other options do not accurately address the primary safety concern associated with polyacrylamide. While some aspects like requiring a protective barrier could be relevant to general lab safety practices, it is the carcinogenic nature of acrylamide that stands out as a key safety issue specifically related to the preparation of the gels. Therefore, understanding the potential health risks associated with acrylamide is crucial for ensuring safe laboratory practices.

5. What is required to form a holoenzyme?

- A. Apoenzyme plus cofactor
- B. Apoenzyme plus substrate
- C. Apoenzyme plus prosthetic group**
- D. Apoenzyme plus reaction product

To form a holoenzyme, it is essential to combine an apoenzyme with a cofactor. A holoenzyme is the active form of an enzyme that includes its non-protein component, which can be a cofactor or a coenzyme. Cofactors can be metal ions or organic molecules, and they are crucial for the enzyme's activity, as they help the enzyme achieve its functional form. The apoenzyme is the protein part of the enzyme that, on its own, is inactive. When it binds to the necessary cofactor, the resulting holoenzyme is capable of catalyzing its specific biochemical reaction. While prosthetic groups, which are tightly bound cofactors, can also contribute to the structure of some holoenzymes, the more general requirement for holoenzyme formation is the combination of an apoenzyme and a cofactor. This distinction is key in understanding enzyme structure and function, as the holoenzyme represents the fully active and functional enzyme complex.

6. How are most enzymes monitored in clinical settings?

- A. By measuring the rate of temperature change
- B. By observing color changes in the reaction
- C. By monitoring the rate of absorbance change**
- D. By assessing pH fluctuations

In clinical settings, most enzymes are monitored by measuring the rate of absorbance change. This method is based on the principle that enzyme-catalyzed reactions often result in the formation or consumption of colored products or substrates that can be quantified using spectrophotometry. As the reaction proceeds, the concentration of the colored species changes, leading to a corresponding change in absorbance at a specific wavelength. By continuously tracking this change in absorbance over time, healthcare professionals can determine the enzyme's activity, which is crucial for diagnosing various medical conditions and assessing metabolic functions. The other methods listed, although they may have applications in specific contexts, are not the primary means by which enzyme activity is routinely monitored in clinical laboratories. For instance, temperature changes are generally not a reliable indicator of enzyme activity due to the complexity of biological reactions and varying temperature responses. Observing color changes, while relevant to certain tests, is less precise than direct absorbance measurements that can quantify changes more accurately. Assessing pH fluctuations is important in enzyme kinetics, but it does not provide a direct measurement of enzyme activity like absorbance changes do. Thus, monitoring the rate of absorbance change is the standard approach in clinical enzyme assays.

7. What is the order of ALP isoenzyme electrophoresis migration from slowest to fastest?

- A. Renal - Intestinal - Bone - Liver - Placental
- B. Renal - Bone - Intestinal - Liver - Placental
- C. Renal - Intestinal - Placental - Bone - Liver
- D. Renal - Intestinal - Bone - Placental - Liver**

The order of alkaline phosphatase (ALP) isoenzyme electrophoresis migration from slowest to fastest is accurately represented in the provided option. To understand this order, it is essential to recognize the different tissue sources of ALP and how their varying molecular weights and structures affect their electrophoretic mobility. ALP isoenzymes originate from various tissues, including the liver, bone, placenta, renal, and intestine, each possessing unique properties. The renal isoenzyme has the lowest mobility due to its larger size and structural characteristics, which results in slower migration through the gel matrix during electrophoresis. Following the renal isoenzyme, the intestinal isoenzyme migrates slightly faster, accompanied by the bone isoenzyme, which also moves at a moderate pace. Next in the order is the placental isoenzyme, which migrates more quickly than the previously mentioned isoenzymes. Finally, the liver isoenzyme, being the most mobile and smallest in size, migrates the fastest. This sequential order of migration reflects their respective physicochemical properties when subjected to electrophoresis, underscoring their varied release profiles from different tissues under physiological and pathological states. The understanding of this migration pattern is essential for interpreting ALP isoenzyme results in clinical contexts.

8. Amylase is classified as what type of enzyme?

- A. Oxidase
- B. Hydrolase**
- C. Transferase
- D. Ligase

Amylase is classified as a hydrolase enzyme, which is responsible for the hydrolysis of glycosidic bonds in carbohydrates. Specifically, amylase catalyzes the breakdown of starch and glycogen into simpler sugars like maltose and glucose by adding water to the molecular bonds. This process of hydrolysis is a key function of hydrolases, which utilize water to cleave chemical bonds. In the context of the other types of enzymes, oxidases are enzymes that facilitate oxidation-reduction reactions, transferases are involved in transferring functional groups between molecules, and ligases link two molecules together, usually requiring energy input such as ATP. These distinctions highlight the specific function and action of amylase within the broader category of enzymes, thereby affirming its classification as a hydrolase.

9. In a lipid panel, which two tests are least affected by fasting?

- A. LDL cholesterol and triglycerides
- B. Total cholesterol and HDL cholesterol**
- C. Triglycerides and HDL cholesterol
- D. LDL cholesterol and total cholesterol

In a lipid panel, total cholesterol and HDL cholesterol levels are generally less affected by fasting compared to other lipid measures like LDL cholesterol and triglycerides. When assessing lipid profiles, fasting is often emphasized primarily due to its significant impact on triglyceride levels, which can be elevated after meals, and LDL calculations that rely on triglyceride levels. Total cholesterol, representing the sum of various types of cholesterol in the blood, tends to remain relatively stable regardless of the fasting state. Similarly, HDL cholesterol, known as "good" cholesterol, is not as influenced by immediate dietary intake. This is because HDL plays a key role in transporting cholesterol to the liver for excretion and is generally less responsive to short-term dietary changes. In contrast, triglycerides are strongly affected by recent food intake and can spike postprandially (after eating). LDL cholesterol, while it is calculated based on the Friedewald formula (which includes triglycerides), can also be influenced by fasting, as an increase in triglycerides would affect the LDL calculation. Thus, total cholesterol and HDL cholesterol are the two tests in a lipid panel that maintain more consistent results regardless of fasting, making the choice of these two tests the most accurate option.

10. Which laboratory phenomenon can affect uric acid testing results significantly?

- A. Specimen hemolysis**
- B. Refrigeration of the sample
- C. Delayed serum separation
- D. Prolonged exposure to light

Uric acid testing can be significantly affected by specimen hemolysis. Hemolysis occurs when red blood cells rupture, releasing intracellular contents, which can interfere with the measurement of various analytes. In the case of uric acid, hemolysis can lead to falsely elevated results due to the release of purines and other metabolites from the broken red blood cells. This alteration in the sample composition affects the accuracy of the uric acid levels detected during analysis. While refrigeration of the sample, delayed serum separation, and prolonged exposure to light can impact other components of laboratory tests, they do not have as profound an effect on uric acid measurements compared to hemolysis. Refrigeration is generally recommended to preserve specimens, and even though delayed serum separation could potentially lead to changes in some analytes over time, it is hemolysis that poses the most immediate and significant risk of contamination and inaccurate results for uric acid testing.

Next Steps

Congratulations on reaching the final section of this guide. You've taken a meaningful step toward passing your certification exam and advancing your career.

As you continue preparing, remember that consistent practice, review, and self-reflection are key to success. Make time to revisit difficult topics, simulate exam conditions, and track your progress along the way.

If you need help, have suggestions, or want to share feedback, we'd love to hear from you. Reach out to our team at hello@examzify.com.

Or visit your dedicated course page for more study tools and resources:

<https://ascpchemistry.examzify.com>

We wish you the very best on your exam journey. You've got this!

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