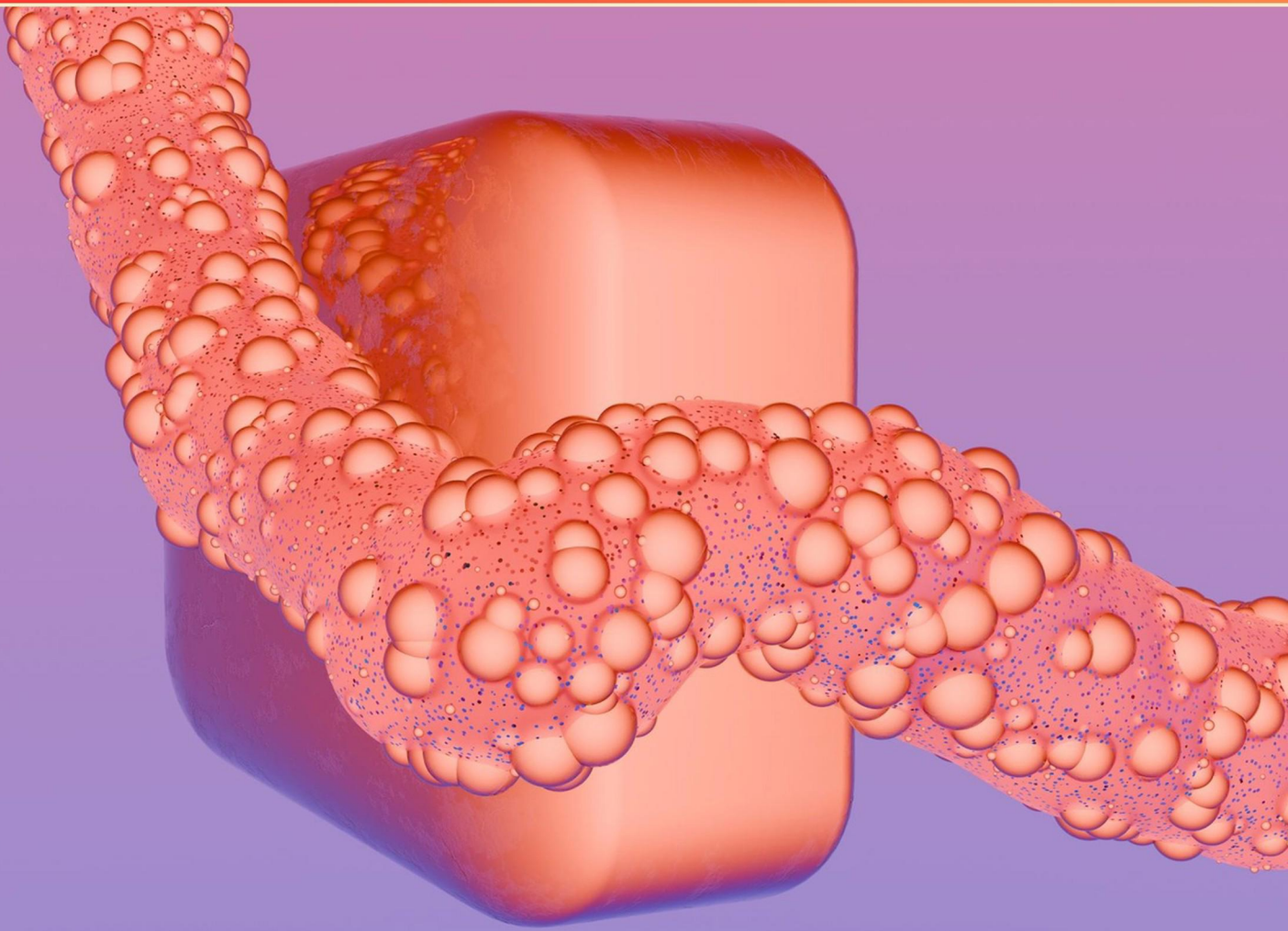


A2 GENETIC CONTROL OF PROTEINS AND CONTROL OF GENE EXPRESSION PRACTICE TEST (SAMPLE)

STUDY GUIDE



**SAMPLE STUDY GUIDE
WITH LIMITED QUESTIONS**

**VISIT US ONLINE FOR
THE FULL VERSION STUDY GUIDE**

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Introduction

Preparing for a certification exam can feel overwhelming, but with the right tools, it becomes an opportunity to build confidence, sharpen your skills, and move one step closer to your goals. At Examzify, we believe that effective exam preparation isn't just about memorization, it's about understanding the material, identifying knowledge gaps, and building the test-taking strategies that lead to success.

This guide was designed to help you do exactly that.

Whether you're preparing for a licensing exam, professional certification, or entry-level qualification, this book offers structured practice to reinforce key concepts. You'll find a wide range of multiple-choice questions, each followed by clear explanations to help you understand not just the right answer, but why it's correct.

The content in this guide is based on real-world exam objectives and aligned with the types of questions and topics commonly found on official tests. It's ideal for learners who want to:

- Practice answering questions under realistic conditions,
- Improve accuracy and speed,
- Review explanations to strengthen weak areas, and
- Approach the exam with greater confidence.

We recommend using this book not as a stand-alone study tool, but alongside other resources like flashcards, textbooks, or hands-on training. For best results, we recommend working through each question, reflecting on the explanation provided, and revisiting the topics that challenge you most.

Remember: successful test preparation isn't about getting every question right the first time, it's about learning from your mistakes and improving over time. Stay focused, trust the process, and know that every page you turn brings you closer to success.

Let's begin.

How to Use This Guide

This guide is designed to help you study more effectively and approach your exam with confidence. Whether you're reviewing for the first time or doing a final refresh, here's how to get the most out of your Examzify study guide:

1. Start with a Diagnostic Review

Skim through the questions to get a sense of what you know and what you need to focus on. Your goal is to identify knowledge gaps early.

2. Study in Short, Focused Sessions

Break your study time into manageable blocks (e.g. 30 – 45 minutes). Review a handful of questions, reflect on the explanations.

3. Learn from the Explanations

After answering a question, always read the explanation, even if you got it right. It reinforces key points, corrects misunderstandings, and teaches subtle distinctions between similar answers.

4. Track Your Progress

Use bookmarks or notes (if reading digitally) to mark difficult questions. Revisit these regularly and track improvements over time.

5. Simulate the Real Exam

Once you're comfortable, try taking a full set of questions without pausing. Set a timer and simulate test-day conditions to build confidence and time management skills.

6. Repeat and Review

Don't just study once, repetition builds retention. Re-attempt questions after a few days and revisit explanations to reinforce learning. Pair this guide with other Examzify tools like flashcards, and digital practice tests to strengthen your preparation across formats.

There's no single right way to study, but consistent, thoughtful effort always wins. Use this guide flexibly, adapt the tips above to fit your pace and learning style. You've got this!

Questions

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- 1. What is the function of a transcriptional repressor?**
 - A. A transcriptional repressor binds RNA to degrade transcripts.
 - B. A transcriptional repressor enhances transcription.
 - C. A transcriptional repressor binds DNA to block transcription, often by blocking RNA polymerase or recruiting co-repressors and chromatin modifiers.
 - D. A transcriptional repressor unwinds DNA.
- 2. How does inhibiting acetylcholinesterase at neuromuscular junctions prevent breathing in the described toxin?**
 - A. Acetylcholine not broken down / stays bound to receptor; Na⁺ ions continue to enter / depolarisation occurs; intercostal muscles stay contracted.
 - B. Acetylcholine is rapidly degraded, preventing receptor activation.
 - C. The toxin blocks Na⁺ channels, preventing action potentials.
 - D. The toxin prevents acetylcholine release from the neuron.
- 3. In the mRNA sequence provided, what is the DNA sequence complementary to the first four bases in this section?**
 - A. GGAC
 - B. CCAG
 - C. CAGG
 - D. CTGA
- 4. Which components participate in determining the order of amino acids during translation?**
 - A. mRNA codon sequence
 - B. tRNA anticodons
 - C. Ribosome
 - D. All of the above
- 5. If a mutation changes a codon but still codes for the same amino acid, what is the effect on the resulting polypeptide?**
 - A. Change in amino acid sequence
 - B. Introduction of a premature stop codon
 - C. No change in amino acid sequence
 - D. Frameshift mutation

- 6. The poly-A tail in mRNA primarily affects which aspects?**
- A. It serves as the coding sequence for amino acids
 - B. It increases mRNA stability and enhances translation efficiency
 - C. It directly participates in splicing
 - D. It signals degradation by exonucleases
- 7. Which statement about post-translational modifications in prokaryotes versus eukaryotes is true?**
- A. Prokaryotes lack many eukaryotic PTMs such as glycosylation.
 - B. Glycosylation is a common PTM in prokaryotes.
 - C. Prokaryotes lack many eukaryotic PTMs; eukaryotic systems provide complex PTMs but are more costly.
 - D. Post-translational modifications do not affect protein function.
- 8. If the second mutation changes valine to alanine in the polypeptide, what could be the effect?**
- A. No change in amino acid sequence
 - B. Valine replaced by alanine in the polypeptide; folding may change
 - C. A stop codon is introduced
 - D. A large insertion occurs
- 9. What is alternative splicing?**
- A. Transcribes RNA in the opposite direction.
 - B. Yields multiple mRNA transcripts from one gene by splicing different combinations of exons.
 - C. Inserts introns into mRNA after transcription.
 - D. Replicates the entire gene to produce multiple copies.
- 10. A protein is encoded by a piece of mRNA 660 nucleotides long. What is the maximum number of amino acids in the protein?**
- A. 180
 - B. 220
 - C. 330
 - D. 660

Answers

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1. C
2. A
3. B
4. D
5. C
6. B
7. C
8. B
9. B
10. B

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Explanations

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1. What is the function of a transcriptional repressor?

- A. A transcriptional repressor binds RNA to degrade transcripts.
- B. A transcriptional repressor enhances transcription.
- C. A transcriptional repressor binds DNA to block transcription, often by blocking RNA polymerase or recruiting co-repressors and chromatin modifiers.**
- D. A transcriptional repressor unwinds DNA.

A transcriptional repressor works by lowering gene expression, mostly by preventing RNA polymerase from transcribing the gene. They usually bind to specific DNA sequences near the promoter or regulatory regions. By sitting on the DNA, they can block RNA polymerase from binding or from moving along the template, stopping transcription. In many cases, they also recruit other proteins that compact chromatin or modify histones, making the DNA less accessible and further reducing transcription. This dual mode—direct interference with the transcription machinery and recruitment of repressive chromatin modifiers—explains how repressors keep genes turned off in different contexts. For example, bacterial repressors like the lac repressor block transcription by occupying the operator, while in eukaryotes, repressors recruit co-repressors and chromatin-modifying enzymes to create a repressive environment. Options that talk about RNA binding to degrade transcripts describe RNA decay rather than transcriptional control, while those claiming enhancement of transcription describe activators. Unwinding DNA is a helicase-like activity, not the typical function of a transcriptional repressor.

2. How does inhibiting acetylcholinesterase at neuromuscular junctions prevent breathing in the described toxin?

- A. Acetylcholine not broken down / stays bound to receptor; Na⁺ ions continue to enter / depolarisation occurs; intercostal muscles stay contracted.**
- B. Acetylcholine is rapidly degraded, preventing receptor activation.
- C. The toxin blocks Na⁺ channels, preventing action potentials.
- D. The toxin prevents acetylcholine release from the neuron.

The key idea is that stopping acetylcholinesterase leaves acetylcholine in the neuromuscular junction, so it keeps binding to nicotinic receptors and continuously activating them. That ongoing receptor activity keeps sodium channels open, causing ongoing depolarization of the muscle end plate. For respiratory muscles, this sustained depolarization means the muscles stay in a contracted state and can't properly relax or coordinate breathing, leading to paralysis of the intercostal muscles and diaphragm and, ultimately, failure to breathe. The other scenarios describe removing acetylcholine, blocking sodium channels, or stopping acetylcholine release, which would prevent contraction rather than cause the dangerous, sustained contraction that blocks breathing.

3. In the mRNA sequence provided, what is the DNA sequence complementary to the first four bases in this section?

- A. GGAC
- B. CCAG**
- C. CAGG
- D. CTGA

Understanding how mRNA bases pair with DNA bases to form a complementary sequence. Each RNA base finds its DNA partner: G pairs with C, U pairs with A, and C pairs with G. To get the DNA sequence complementary to the first four bases of the mRNA, substitute each base with its DNA partner: G becomes C, the second G becomes C, U becomes A, and C becomes G. Put together, that gives CCAG. This is why CCAG fits—the four mRNA bases GGUC pair with CCAG on the DNA template. The other sequences would arise from different RNA bases in those positions, so they wouldn't match the given mRNA segment.

4. Which components participate in determining the order of amino acids during translation?

- A. mRNA codon sequence
- B. tRNA anticodons
- C. Ribosome
- D. All of the above**

The order of amino acids during translation is set by the combination of the mRNA codon sequence, the tRNA anticodons that pair with those codons, and the ribosome that coordinates decoding and peptide bond formation. The mRNA carries the genetic code in codons, each specifying which amino acid should be added next. tRNA molecules deliver the appropriate amino acids, with anticodons that match the codons to ensure the correct amino acid is chosen for each step. The ribosome brings the mRNA and tRNA together, catalyzes the formation of peptide bonds, and moves along the mRNA to maintain the proper reading frame, ensuring the sequence of amino acids matches the codon order. Because all three components are essential for translating the code into a polypeptide, they collectively determine the final sequence of amino acids.

5. If a mutation changes a codon but still codes for the same amino acid, what is the effect on the resulting polypeptide?

- A. Change in amino acid sequence
- B. Introduction of a premature stop codon
- C. No change in amino acid sequence**
- D. Frameshift mutation

When a codon changes but still encodes the same amino acid, the amino acid sequence of the protein remains unchanged. This is because multiple codons can specify the same amino acid due to the degeneracy of the genetic code. Such a mutation is called a silent (synonymous) mutation, and the primary structure of the polypeptide is the same, so the protein's overall properties are usually preserved. In rare cases, there might be subtle effects on translation efficiency or folding due to codon usage, but the actual amino acid sequence doesn't differ. If a different amino acid were incorporated, the protein's sequence would change. If a premature stop codon appeared, the polypeptide would be truncated. If nucleotides were inserted or deleted not in multiples of three, the reading frame would shift, altering downstream amino acids (a frameshift).

6. The poly-A tail in mRNA primarily affects which aspects?

- A. It serves as the coding sequence for amino acids
- B. It increases mRNA stability and enhances translation efficiency**
- C. It directly participates in splicing
- D. It signals degradation by exonucleases

The poly-A tail mainly increases mRNA stability and translation efficiency. After transcription, a stretch of adenines is added to the 3' end and binds poly(A)-binding proteins. These proteins couple with the cap-binding complex to circularize the mRNA, which makes ribosome recruitment and reinitiation more efficient, boosting how well the message is translated. The tail also shields the mRNA from 3' to 5' exonucleases, extending its lifespan so more protein can be produced over time. It doesn't encode amino acids, and splicing is carried out before polyadenylation, so the tail isn't directly involved in splicing. While shortening of the tail can lead to decay, its primary role is enhancing stability and translation, not signaling degradation.

7. Which statement about post-translational modifications in prokaryotes versus eukaryotes is true?

- A. Prokaryotes lack many eukaryotic PTMs such as glycosylation.
- B. Glycosylation is a common PTM in prokaryotes.
- C. Prokaryotes lack many eukaryotic PTMs; eukaryotic systems provide complex PTMs but are more costly.**
- D. Post-translational modifications do not affect protein function.

Post-translational modifications after protein synthesis expand function and regulation; the main difference between prokaryotes and eukaryotes is the breadth and complexity of these modifications. Prokaryotes do perform some PTMs, but they generally lack the wide, highly regulated set found in eukaryotic cells. Eukaryotes can implement intricate modifications such as elaborate glycosylation patterns, extensive phosphorylation networks, ubiquitination, and other refinements that fine-tune activity, stability, and interactions. Because of this greater complexity, using eukaryotic expression systems to obtain proteins with these PTMs is more costly due to slower growth, more demanding growth conditions, and more complex processing. Although a few prokaryotes can perform certain PTMs like glycosylation or phosphorylation, they are not as prevalent or sophisticated as in eukaryotes, so the statement that captures the overall truth is that prokaryotes lack many eukaryotic PTMs while eukaryotic systems provide complex PTMs at higher cost. And of course, PTMs do affect protein function; ignoring them would miss a core aspect of how proteins are regulated.

8. If the second mutation changes valine to alanine in the polypeptide, what could be the effect?

- A. No change in amino acid sequence
- B. Valine replaced by alanine in the polypeptide; folding may change**
- C. A stop codon is introduced
- D. A large insertion occurs

The main concept is what happens when a single amino acid in a protein is changed. This is a missense mutation, where one amino acid is substituted for another in the polypeptide chain. Replacing valine with alanine alters the side chain at that position. Valine has a larger, branched hydrophobic side chain, while alanine is smaller. This change can affect how the protein folds because folding depends on how side chains pack together and interact in the interior and at interfaces. Even a smaller, similar nonpolar substitution can tweak packing enough to alter local structure, stability, or function, so the folding could change. The other possibilities describe different types of mutations that aren't occurring here: a silent mutation would leave the amino acid unchanged, a stop codon introduction would truncate the protein (nonsense mutation), and a large insertion would disrupt the reading frame (frameshift).

9. What is alternative splicing?

- A. Transcribes RNA in the opposite direction.
- B. Yields multiple mRNA transcripts from one gene by splicing different combinations of exons.**
- C. Inserts introns into mRNA after transcription.
- D. Replicates the entire gene to produce multiple copies.

Alternative splicing is the process by which a single gene can generate multiple mRNA transcripts by joining exons in different combinations during RNA processing. After transcription, the pre-mRNA carries exons and introns, and the spliceosome can select different splice sites or skip certain exons. This yields distinct mature mRNAs from the same gene, leading to different protein variants. It's not about transcribing RNA in the opposite direction, not about inserting introns after transcription (introns are removed), and not about duplicating the gene.

10. A protein is encoded by a piece of mRNA 660 nucleotides long. What is the maximum number of amino acids in the protein?

- A. 180
- B. 220**
- C. 330
- D. 660

Genes are read in codons of three nucleotides, and each codon specifies one amino acid (with stop codons signaling termination). Therefore, the maximum number of amino acids that can be encoded by an mRNA segment is the total nucleotides divided by 3, rounding down if needed. For 660 nucleotides, that's $660 \div 3 = 220$ codons that could code amino acids. The protein could reach 220 amino acids in the maximum-case scenario where no stop codon occurs within the coding region. (If a stop codon were present, the count would drop by one, but the question asks for the maximum, so 220 is the best answer.)

Next Steps

Congratulations on reaching the final section of this guide. You've taken a meaningful step toward passing your certification exam and advancing your career.

As you continue preparing, remember that consistent practice, review, and self-reflection are key to success. Make time to revisit difficult topics, simulate exam conditions, and track your progress along the way.

If you need help, have suggestions, or want to share feedback, we'd love to hear from you. Reach out to our team at hello@examzify.com.

Or visit your dedicated course page for more study tools and resources:

<https://a2genecontofproteinsandgeneexpression.examzify.com>

We wish you the very best on your exam journey. You've got this!

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